

## A Case of Localized Amyloid Angiopathy Following Aneurysmal Subarachnoid Hemorrhage - Strong Evidence of Dysfunction of the Glymphatic System

Forshing Lui<sup>1\*</sup>, Ning Zhong<sup>2</sup>, Kaho Wong<sup>3</sup>, Stella Knowlton<sup>1</sup>, Jessa Alcaide<sup>1</sup> and Michael Ysit<sup>1</sup>

<sup>1</sup>California Northstate University College of Medicine, Elk Grove, CA, USA

<sup>2</sup>Kaiser Permanente Medical Center, Sacramento, CA, USA

<sup>3</sup>Kaiser Permanente Medical Center, Roseville, CA, USA

**\*Corresponding Author:** Forshing Lui, California Northstate University College of Medicine, Elk Grove, CA, USA.

**Received:** October 21, 2021

**Published:** November 12, 2021

© All rights are reserved by **Forshing Lui, et al.**

### Abstract

There are no lymphatics within the brain like other organs in our body with traditional anatomical understanding until Iliff and Nedergaard demonstrated a system of cerebrospinal and brain interstitial fluid flow systems that appears to be similar to the lymphatic system in rodents. They created the term "glymphatics". More recent studies by using MRI tracer studies in humans demonstrated evidences of a similar glymphatic system in humans. It is logical to believe that such a system functions as an important waste clearance system in health and disease. However, the function of such a system has been questioned by more recent studies. There has never been any real human case to confirm the pathophysiologically the dysfunction of such a system as the reason for the underlying disease. Furthermore, there has never been any explanation how beta-amyloid synthesized in the brain can be transported retrogradely to the small brain arteries and arterioles in amyloid angiopathy.

We report a case of relatively rapid development of amyloid angiopathy over a few years after an acute subarachnoid hemorrhage secondary to rupture of the posterior communicating aneurysm. The amyloid angiopathy is relatively localized over the site of subarachnoid hemorrhage. There are also associated local changes and amyloid deposition in the brain parenchyma.

We believe this case evidently and adequately demonstrated the function and dysfunction of the glymphatic system in waste clearance and movement of solutes in the brain parenchyma. It also demonstrated how a-beta amyloid is synthesized in the brain parenchyma and transported retrogradely towards and got deposited in the small cerebral arteries in amyloid angiopathy.

**Keywords:** Glymphatics; A-beta Amyloid; Amyloid Angiopathy; Proteinopathy

### Abbreviations

CSF: Cerebrospinal Fluid; ISF: Interstitial Fluid; AD: Alzheimer's Disease; A $\beta$ : A-beta; AQP4: Aquaporin-4; MOCA: Montreal Cognitive Assessment Test; CAA: Cerebral Amyloid Angiopathy

### Introduction

The existence of a brain lymphatic system similar to systemic lymphatics is unknown to most neurologists and many neuroscientists. The present day, medical and neuroscience students learn

the traditional concept of brain CSF production by the choroid plexus, circulating through the 4 ventricles and the subarachnoid space and returning to the systemic circulation through the arachnoid granulations. The modern understanding of the CSF flow dynamics has been revolutionized by the work done by Iliff and Nedergaard, et al. [1,2] on rodents. They created the term "glymphatics" to describe the flow of cerebrospinal fluid (CSF) from the perivascular space (Virchow Robbin, VR space) accompanying the small cortical arteries or arterioles into the brain interstitial fluid (ISF) and

drained out of the system into the deep cervical lymphatics. The efflux of CSF into the brain interstitial fluid depends heavily on the function and integrity of the astroglial water channels (Aquaporin-4, AQP4). Impaired CSF-ISF movement was demonstrated in AQP4 knock-out mice [3]. More recent human studies by using MRI with CSF tracer [4,8-11] demonstrated a similar system and its flow pattern in normal and some disease states. Taoka and Naganawa [8] used the term “CNS interstitial fluidopathy” to define neurological disorders caused by impaired CNS interstitial fluid dynamics. It was also postulated that the water channels (AQP4) in the astroglial foot processes play an important part in the exchange of fluid and solutes between the CSF and the ISF. The glymphatic system provides an important waste clearance system for the brain to remove large molecular solutes and wastes such as amyloid beta (A $\beta$ ) and tau proteins. Impairment of this clearance system may play an important part in the pathogenesis of various neurodegenerative disorders including Alzheimer’s disease, age-related changes, idiopathic normal pressure hydrocephalus, and traumatic brain injury [4,5]. However, different viewpoints arose recently questioning the significance of this glymphatic clearance system [6,7].

### The case

Our patient is a 59-year-old lady with known history of well-controlled hypertension, hyperlipidemia, and chronic smoking. She presented with an acute subarachnoid hemorrhage at the age of 53 (February 2016) due to rupture of her right PCoM aneurysm (CT, Figure 1). Her aneurysm was clipped surgically. She was discharged home with quite good functional recovery. Her main deficit was mild clumsiness with her gait and motor activities.

**Figure 1:** Acute SAH due to ruptured R PCoM aneurysm(arrow) (February 2016).

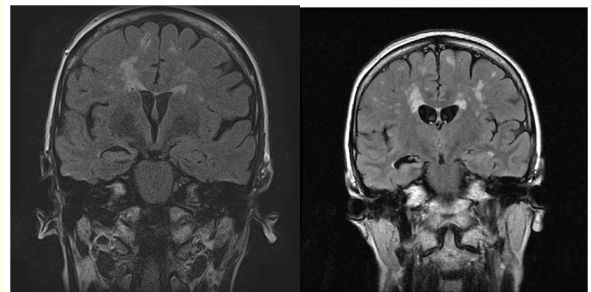
She presented again to our neurological service in 2018 with recurrent complex partial seizures with secondary generalization.

Her EEG was reported as showing right temporoparietal periodic epileptiform discharge. On subsequent follow-up EEG, there were left fronto-temporal epileptiform discharges. Her seizures were managed with Lacosamide and Valproate.

Since 2020, her family noticed some problems with memory and attention. Examination in April 2021 reviewed a MOCA score of 28/30 with one point miss each in visuospatial function and delayed recall. Her NACC Functional Assessment (Functional Activities Questionnaire (FAQ) was 11/27 (Abnormal > 9/30). The clinical diagnosis was mild cognitive impairment.

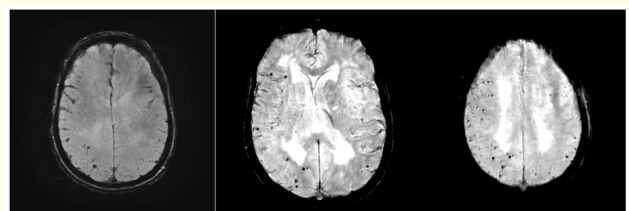
From 2018 until 2021, the patient had a repeated brain MRI performed. The most significant findings include:

- Severe progressive atrophy of her right hippocampus with relatively normal size of her left hippocampus (Figure 2).



**Figure 2:** MRI - FLAIR sequence: Progressive R hippocampal atrophy (August 2018) and (March 2021).

- Progressive development of microhemorrhages as shown in serial GRE/SWI MRI. These are lobar microhemorrhages sparing deep hemisphere structures, most compatible with probable amyloid angiopathy according to the modified Boston Criteria. The diagnosis is further supported by the positive Amyloid PET scan (Figures 3 and 4).



**Figure 3:** Progressive development of R>L hemisphere amyloid angiopathy (GRE and SWI sequences).

**Figure 4:** Diffusely positive amyloid deposition in Amyloid PET Scan: more prominent on the R hemisphere.

- The microhemorrhages increased in number from 2018 to 2021. The number is consistently and significantly higher affecting her right cerebral hemisphere (Figure 3).
- Her recent amyloid PET scan showed diffuse cerebral amyloid deposition, slightly more on her right hemisphere (Figure 4).

Our case is unique and there has not been any similar case reported in the literature. It illustrated several important clinicopathological changes which may help in the understanding of neurodegenerative disorders as a result of impaired waste (protein) clearance from the brain as follows:

- The patient was neurologically normal clinically before she suffered from an acute subarachnoid hemorrhage secondary to rupture of a right posterior communicating artery aneurysm. She might already have some early Alzheimer's disease pathology as evidenced by the widespread and diffuse amyloid deposition demonstrated in her recent amyloid PET study.
- She developed apparent progressive clinicopathological changes in her brain after her acute subarachnoid hemorrhage. These include progressive cognitive impairment, progressive right hemisphere worse than left hemisphere amyloid angiopathy (modified Boston criteria), progressive right hippocampal atrophy. These can all be explained by dysfunction of the glymphatic clearance system as a result of the subarachnoid hemorrhage as discussed below.

## Discussion

Amyloid beta (A $\beta$ ) peptide accumulation in the brain is one of the two main pathological hallmarks of Alzheimer's disease (AD). A $\beta$  is deposited in extracellular neurites in the brain forming amyloid neuritic plaques, especially in areas including the temporoparietal neocortex and the hippocampus associated with early atrophy of the hippocampus in patients with AD. Hardy and Higgins reviewed molecular and biological evidence in molecular genetics suggesting A $\beta$  is central to the etiology of AD [12,13]. The other pathological hallmark is the accumulation of hyperphosphorylated tau (pTau) protein inside the neurons as neurofibrillary tangles. Pathological A $\beta$  is produced by sequential proteolytic cleavage of the transmembrane protein, amyloid precursor protein (APP) by  $\beta$ - and  $\gamma$ -secretase enzymes [14]. Hardy and Higgins [12] postulated the amyloid cascade hypothesis in 1992 in the pathogenetic mechanism of AD. Subsequent studies, however, failed to show the correlation between plaque load and cognitive decline in patients with AD [15]. Nevertheless, the presence of A $\beta$  neuritic plaques is still the most important pathological hallmark to define the disease. Imaging of A $\beta$  such as amyloid PET scan is also used to identify underlying brain pathology as AD in patients with dementia or in clinical research.

The main pathological substrates in the brain and brain arteries are A $\beta$ 1-42 predominantly in neocortical and hippocampal neurites and the smaller and more soluble A $\beta$ 1-40 in the arterial walls of small cerebral arteries and arterioles as amyloid angiopathy (CAA) [16]. It is interesting to note that CAA is uncommon in deep hemispheric structures including basal ganglia, thalamus, and brainstem. It is easily understandable that excessive synthesis of A $\beta$ 1-42 will lead to deposition in neurites especially in genetic and familial forms of AD such as cases due to presenilin-1 mutation, Down's syndrome, and Apolipoprotein E4 genotypes. It will be difficult to visualize how A $\beta$ 1-40 which is also synthesized in the brain, can be "transported" retrogradely to the cerebral arteries and deposited in the walls of these arteries and arterioles resulting in subsequent complications of hemorrhage and ischemia. In the detailed review of A $\beta$  [17], Chen, *et al.* mentioned its clearance across the blood-brain barrier (BBB) as one of the mechanisms of removal or clearance. It will be easier to understand its clearance down the pressure gradient towards the venous system than on the pressure gradient towards the arterial system.

Our case illustrates many unanswered questions about A $\beta$  clearance and its increased deposition inside the cerebral neocor-

tex and small arteries and arterioles. First, we need to review a more recently discovered cerebral waste clearance system involving the cerebrospinal fluid (CSF) and brain interstitial fluid (ISF) called the “glymphatics” system.

There is about 150 mL of CSF in an average adult. The 24 hours turnover is about 500 mL. The main known function of the CSF is to provide buoyancy for our brain. It is easily understood that there should be more important functions to explain the high CSF turnover. The logical reason should include metabolic supply and waste clearance for our metabolically active brain. The traditional view begins with its formation by the choroid plexus. It circulates through the 4 ventricles and is finally reabsorbed by the subarachnoid granulations. This concept has been challenged more recently both in CSF production and CSF removal [18,19]. For CSF production, a more recent concept supported by good scientific evidence is that a large volume of CSF is produced directly from the capillaries in the brain secondary to hydrostatic pressure, rather than strictly from the choroid plexus [20,21].

The removal of CSF solely by arachnoid granulations has been under stronger scrutiny by many studies over the past decade or so. Tracer studies of CSF flow [22,23] showed drainage through the lymphatics of the cribriform plate, cranial, and spinal nerve sheaths. The most significant contribution to our modern understanding of CSF drainage and brain waste clearance, and the existence of a brain structure similar to systemic lymphatics is the concept of “glymphatics” by Iliff and Nedergaard [24].

By using fluorescent CSF tracers on rodents, Iliff, *et al.* were able to demonstrate a large volume of CSF in the subarachnoid space rapidly entering the brain parenchyma along the perivascular spaces (Virchow-Robin or VR space) of the cortical penetrating arteries. The perivascular CSF space and the brain interstitial fluid (ISF) space are separated by the foot processes of astrocytes which have abundant water channels (aquaporin-4, AQP4), which are polarized and allow only unidirectional movement of fluid. At this point, the CSF in the Virchow-Robin space was driven out to the brain ISF space by arterial pulsation and convective bulk flow. This CSF influx occurs diffusely and uniformly throughout all cortical deep penetrating arteries centripetally towards the perivenous spaces of the deep draining veins with drainage more restricted towards several large draining veins. Fluorescent tracers from deep nuclear structures such as the striatum and thalami are drained

medially to the perivenous spaces of the great vein of Galen and the internal cerebral veins. The CSF-ISF flow enters the perivenous space again through the AQP4 channels and is finally drained from the brain through the deep cervical lymphatics.

More recent studies [10,11,23,25,26] by using MRI with gadobutrol CSF tracer demonstrated the glymphatic system in humans quite convincingly. Intrathecally administered CSF tracer enters the brain from cortical surfaces alongside the periarterial spaces and moves centripetally towards the deep structures. Clearance of tracer is delayed in aging patients and patients with dementia and patients with normal pressure hydrocephalus [27,28].

The glymphatic CSF and ISF flow alters with different physiological and pathological conditions. Natural sleep is associated with enhanced glymphatic flow and clearance [29,30]. Impaired glymphatic flow is shown in old age, in idiopathic normal pressure hydrocephalus, and traumatic brain injury [27,28].

The prevalence of CAA increases with age. Associations between CAA APOE4 genotype and Alzheimer’s disease neuropathological changes (ADNC) has been documented with multiple studies [31-33]. 78 - 98% of patients with ADNC were estimated to have CAA. On the other hand, only 25% of patients with CAA have ADNC. CAA is caused by deposition of the more soluble A $\beta$ 1-40 whereas amyloid plaque consists mainly of the less soluble A $\beta$ 1-42. A $\beta$ 1-42 is synthesized inside the brain from the more soluble oligomers with the pleated sheet structure in neuritic plaques. Pathological condition related, as in Alzheimer’s disease, or age-related increase in A $\beta$  production in the brain is considered the cause of the deposition in the neurites or cerebral arteries. The A $\beta$ 1-40 is also produced in the brain. How it ends up in the cerebral arteries retrogradely with the CSF-ISF flow has not been studied. With the understanding of the CSF-ISF glymphatic flow system, there will be A $\beta$ 1-42 deposition if it is not adequately cleared from the brain. Our case demonstrated an impairment of glymphatic flow on the arterial side due to fibrin deposition and other mechanisms as a result of her acute subarachnoid hemorrhage. The AQP4 channels are expected to be damaged, and the polarity of flow may also be altered. In addition, the centripetal glymphatic flow from the cortical VR space to the deep venous perivenous spaces may be altered from a fluid dynamics standpoint. If there is impaired glymphatic inflow from the cortical areas affected by pathological changes in the VR CSF space and astroglial AQP4 channels secondary to the acute subarach-

noid hemorrhage, the CSF-ISF may be directed towards these areas. When the polarity of the AQP4 channels is damaged, it will be easy to understand how the A $\beta$ 1-40 may be carried along with the ISF in

a retrograde manner towards the cerebral arteries and becomes deposited in the arterial wall with the consequences (Figure 5).

**Figure 5:** Bulk CSF-ISF flow in normal and abnormal in our patient after her acute aneurysmal subarachnoid hemorrhage: amyloid AB42 deposition in neurites and AB40 in cerebral arteries - dysfunction of the glymphatic system.

Rowe., *et al.* [34] showed evidence suggesting that A $\beta$  deposition precedes the diagnosis of Alzheimer's disease by about 15 years. The amyloid PET scan of our patient performed in 2021 showed the presence of diffuse amyloid deposition in both hemispheres. It will be logical to assume that our patient started A $\beta$  deposition in her brain before she developed her acute subarachnoid hemorrhage. Her amyloid PET scan did show some asymmetry with more deposition in her right hemisphere, another evidence to suggest the impaired glymphatic function may have accelerated the A $\beta$  deposition in the cerebral neurites.

## Conclusion

The glymphatic system involving the CSF, brain ISF, and the astrocytic foot processes lined with AQP4 channels should be considered as an important waste clearance system of our brain. It was underrecognized until very recently. It may play an important part in many neurodegenerative disorders including Alzheimer's disease. Our unique case of the progressive development of amyloid angiopathy and asymmetric hippocampal atrophy following an acute subarachnoid hemorrhage due to rupture of her right pos-

terior communicating artery aneurysm demonstrates strong evidence suggesting dysfunction of the glymphatic clearance system. Future therapeutic approaches may focus more on improving this clearance system for many neurodegenerative disorders.

## Author Contributions

- Forshing Lui: Preparation and drafting of the whole manuscript.
- Ning Zhong: Provision, management and discussions about clinical cases and review of the manuscript.
- Kaho Wong: Provision, management and discussions about clinical cases and review of the manuscript.
- Stella Knowlton, Jessa Alcaide, and Michael Ysit: Preparation of the figure and review of the manuscript.

## Funding

None.

## Conflict of Interest

All authors have NO conflict of interest exist.

## Bibliography

1. Iliff JJ Wang M., *et al.* "A paravascular pathway facilitates CSF flow through the brain parenchyma and the clearance of interstitial solutes, including amyloid  $\beta$ ". *Science Translational Medicine* 4.147 (2012): 147ra111.
2. Iliff J and Simon M. "The glymphatic system supports convective exchange of cerebrospinal fluid and brain interstitial fluid that is mediated by perivascular aquaporin-4". *The Journal of Physiology* 597.17 (2019): 4417.
3. Mestre H., *et al.* "Aquaporin-4-dependent glymphatic solute transport in the rodent brain". *Elife* 7 (2018): e40070.
4. Davoodi-Bojd E., *et al.* "Modeling glymphatic system of the brain using MRI". *Neuroimage* 188 (2019): 616-627.
5. Jessen NA., *et al.* "The glymphatic system: a beginner's guide". *Neurochemical Research* 40.12 (2015): 2583-2599.
6. Smith AJ., *et al.* "Test of the 'glymphatic' hypothesis demonstrates diffusive and aquaporin-4-independent solute transport in rodent brain parenchyma". *Elife* 6 (2017): e27679.
7. Smith AJ and Verkman AS. "The "glymphatic" mechanism for solute clearance in Alzheimer's disease: game changer or unproven speculation?" *The FASEB Journal* 32.2 (2018): 543-551.
8. Ringstad G., *et al.* "Brain-wide glymphatic enhancement and clearance in humans assessed with MRI". *JCI Insight* 3.13 (2018).
9. Taoka T and Naganawa S. "Imaging for central nervous system (CNS) interstitial fluidopathy: disorders with impaired interstitial fluid dynamics". *Japanese Journal of Radiology* 39.1 (2021): 1-4.
10. Watts R., *et al.* "Measuring glymphatic flow in man using quantitative contrast-enhanced MRI". *American Journal of Neuroradiology* 40.4 (2019): 648-651.
11. Eide PK., *et al.* "Magnetic resonance imaging provides evidence of glymphatic drainage from human brain to cervical lymph nodes". *Scientific Reports* 8.1 (2018): 1-10.
12. Hardy JA and Higgins GA. "Alzheimer's disease: the amyloid cascade hypothesis". *Science* 256.5054 (1992): 184-186.
13. Tanzi RE. "The synaptic A $\beta$  hypothesis of Alzheimer disease". *Nature Neuroscience* 8.8 (2005): 977-979.
14. Chen GF., *et al.* "Amyloid beta: structure, biology and structure-based therapeutic development". *Acta Pharmacologica Sinica* 38.9 (2017): 1205-1235.
15. Engler H., *et al.* "Two-year follow-up of amyloid deposition in patients with Alzheimer's disease". *Brain* 129.11 (2006): 2856-2866.
16. Yamada M. "Cerebral amyloid angiopathy: emerging concepts". *Journal of Stroke* 17.1 (2015): 17.
17. Chen GF., *et al.* "Amyloid beta: structure, biology and structure-based therapeutic development". *Acta Pharmacologica Sinica* 38.9 (2017): 1205-1235.
18. Plog BA and Nedergaard M. "The glymphatic system in central nervous system health and disease: past, present, and future". *Annual Review of Pathology: Mechanisms of Disease* 13 (2018): 379-394.
19. Taoka T and Naganawa S. "Neurofluid dynamics and the glymphatic system: a neuroimaging perspective". *Korean Journal of Radiology* 21.11 (2020): 1199.
20. Orešković D and Klarica M. "The formation of cerebrospinal fluid: nearly a hundred years of interpretations and misinterpretations". *Brain Research Reviews* 64.2 (2010): 241-262.
21. Nakada T and Kwee IL. "Fluid dynamics inside the brain barrier: current concept of interstitial flow, glymphatic flow, and cerebrospinal fluid circulation in the brain". *The Neuroscientist* 25.2 (2019): 155-166.
22. Watts R., *et al.* "Measuring glymphatic flow in man using quantitative contrast-enhanced MRI". *American Journal of Neuroradiology* 40.4 (2019): 648-651.
23. Eide PK., *et al.* "Magnetic resonance imaging provides evidence of glymphatic drainage from human brain to cervical lymph nodes". *Scientific Reports* 8.1 (2018): 1-10.
24. Iliff JJ and Nedergaard M. "Is there a cerebral lymphatic system?" *Stroke* 44.6-1 (2013): S93-S95.
25. Kaur J., *et al.* "Magnetic resonance imaging and modeling of the glymphatic system". *Diagnostics* 10.6 (2020): 344.
26. Yang L., *et al.* "Evaluating glymphatic pathway function utilizing clinically relevant intrathecal infusion of CSF tracer". *Journal of Translational Medicine* 11.1 (2013): 1-9.
27. Kress BT., *et al.* "Impairment of paravascular clearance pathways in the aging brain". *Annals of Neurology* 76.6 (2014): 845-861.
28. Reeves BC., *et al.* "Glymphatic system impairment in Alzheimer's disease and idiopathic normal pressure hydrocephalus". *Trends in Molecular Medicine* 26.3 (2020): 285-295.

29. Xie L., *et al.* "Sleep drives metabolite clearance from the adult brain". *Science* 342.6156 (2013): 373-377.
30. Hauglund NL., *et al.* "Cleaning the sleeping brain—the potential restorative function of the glymphatic system". *Current Opinion in Physiology* 15 (2020): 1-6.
31. Biffi A and Greenberg SM. "Cerebral amyloid angiopathy: a systematic review". *Journal of Clinical Neurology* 7.1 (2011): 1-9.
32. Greenberg SM and Charidimou A. "Diagnosis of cerebral amyloid angiopathy: evolution of the Boston criteria". *Stroke* 49.2 (2018): 491-497.
33. Brenowitz WD., *et al.* "Cerebral amyloid angiopathy and its co-occurrence with Alzheimer's disease and other cerebrovascular neuropathologic changes". *Neurobiology of Aging* 36.10 (2015): 2702-2708.
34. Rowe CC., *et al.* "Amyloid imaging results from the Australian Imaging, Biomarkers and Lifestyle (AIBL) study of aging". *Neurobiology of Aging* 31.8 (2010): 1275-1283.

**Volume 4 Issue 12 December 2021**

**© All rights are reserved by Forshing Lui., *et al.***