

Volume 4 Issue 11 November 2023

# Distinct Phenotypic Patterns of *ABCA4* Associated Retinal Disease in a Unique Saudi Consanguineous Cohort

#### Basamat AlMoallem<sup>1,2\*</sup>

<sup>1</sup>Department of Ophthalmology, College of Medicine, King Saud University, Riyadh, Saudi Arabia

<sup>2</sup>Department of Ophthalmology, King Saud University Medical City (KSUMC), Riyadh, Saudi Arabia

\*Corresponding Author: Basamat AlMoallem, Department of Ophthalmology, College of Medicine, King Saud University, King Abdul-Aziz University Hospital, King Abdul Aziz Rd, Al Malaz, Riyadh, Saudi Arabia. Received: September 28, 2023 Published: October 23, 2023 © All rights are reserved by Basamat AlMoallem.

#### Abstract

**Purpose:** This study aimed to establish phenotype- genotype correlations in a unique cohort of thirteen patients from eight unrelated consanguineous Saudi families with variable inherited retinal phenotypes caused by *ABCA4* mutations.

**Methods:** A detailed ophthalmological assessment followed by next-generation sequencing (NGS) screening using vision panel was performed in the affected patients. Reference sequence (*ABCA4*): NM\_000350.

**Results:** Clinically, all patients had reduced visual acuity and nystagmus documented in early childhood. A considerable phenotypic variability of *ABCA4* associated disease (AAD) was observed including: Stargardt disease in 2/13 (15%), Retinitis Pigmentosa in 5/13 (39%) and Cone- Rod dystrophy in 6/13 (46%) and). Genetically; potentially eight distinct homozygous ABCA4 mutations were identified in all the studied patients (100%), five of which are novel.

**Conclusion:** Our study represents a unique cohort of patients originating from consanguineous families with variable retinal phenotypes caused by known and novel recessive *ABCA4* mutations. This study expands the phenotypic – genotypic spectrum of AAD and provides an accurate genetic diagnosis that paves the gates for the opportunity of gene-based therapies in AAD within the scope of precision medicine

Keywords: ABCA4; Stargardt Disease; Mutation; Cone-Rod Dystrophy; Rapid-Onset Chorioretinopathy

### Introduction

Inherited retinal diseases (IRDs) is a heterogeneous group of visually debilitating diseases that are caused by mutations in critical genes to retinal function [1]. Out of this, *ABCA4* associated disease (AAD) is a unique spectrum of IRDs that has been linked to mutations in the *ABCA4* (encoding the transmembrane protein ABCA4 (ATP-binding cassette (ABC), subfamily A, member 4, MIM 601691) [2].

*ABCA4* was found to be highly expressed in the outer-segment of rod and cone photoreceptors and plays a pivotal role in the visual cycle by preventing premature retinal pigmented epithelium (RPE) cell death and photoreceptors degradation [3]. Although, week expressions of *ABCA4* was detected in fibroblasts, lymphocytes, keratinocytes, kidney and brain but no clear functional correlation was generated so far.

Up to date, biallelic mutations in *ABCA4* can lead to a wide spectrum of AAD phenotypes including: Stargardt disease 1 (STGD1), cone-rod dystrophy (CRD), a typical retinitis pigmentosa (RP), fundus flavimaculatus (FFM), rapid onset Chorioretinopathy (ROC) and generalized choriocapillaris atrophy (GCCD). On the other hand, these distinct phenotypes are based on age of onset, disease progression and the spectrum of retinal degeneration [1].

Here, we aimed to clinically and genetically and characterize a unique cohort of 13 individuals from 8 unrelated consanguineous

**Citation:** Basamat AlMoallem. "Distinct Phenotypic Patterns of *ABCA4* Associated Retinal Disease in a Unique Saudi Consanguineous Cohort". *Acta Scientific Clinical Case Reports* 4.11 (2023): 30-35.

families with autosomal recessive AAD, to unequivocally confirm the diagnosis and to establish phenotype- genotype correlations.

#### Methods

#### Patients and clinical assessment

This study was conducted following the IRB-approved written informed consent by King Saud University Institutional Review Board, (KSU- IRB Project NO. E-23-7817). Thirteen patients out of eight unrelated consanguineous Saudi families with variable inherited retinal phenotypes (simplex = 5, complex = 8) were recruited from the ophthalmic genetics clinic at Department of Ophthalmology, College of Medicine, King Saud University (KSU), Riyadh, Saudi Arabia. These patients underwent a detailed ophthalmological examination including best corrected visual acuity (BCVA), dilated fundus examination, retinal fundus imaging, fundus autofluorescence (FAF) imaging and optical coherence tomography (OCT) scans. Full-field electroretinography (ERG) was performed according to the International Society for Clinical Electrophysiology of Vision (ISCEV) standards [4,5]. Genomic DNA was extracted from EDTA blood using standard procedures.

# Mutation screening by next-generation multiplexing -Vision panel assay

Genomic DNA was extracted from EDTA blood using standard procedures. All index patients were initially run on Vision panel as a first-tier test. A panel of 322 genes known to be mutated in various ophthalmic genetic diseases including those involving in inherited retinal conditions was designed as previously described [6]. Genes were amplified using the AmpliSeq HiFi mix and proprietary primer design (Thermo Fisher, Carlsbad, CA), followed by sequencing on the Ion Proton platform following the manufacturer's protocol.

#### Variant interpretation

Variant classification as pathogenic, likely pathogenic or variants of unknown significance (VOUS) was performed following ACMG guidelines [7]. The variant filtering pipeline was applied using the unique Saudi Human Genome in addition to other general genomic databases including dbSNP build 145 (http://www. ncbi.nlm.nih.gov/SNP/), ExAC (http://exac.broadinstitute.org) and gnomAD (http://gnomad.broadinstitute.org). Moreover, *in silico* prediction tools were used: Align GVGD, Sorting Intolerant From Tolerant [SIFT], MutationTaster, and PolyPhen-2, Grantham score calculation, conservation. Mutation nomenclature uses numbering with the A of the initiation codon ATG as +1 (http://varnomen. hgvs.org/) based on the following RefSeqs: NM\_000350 (*ABCA4*).

#### Results

#### **Clinical assessment**

A total of 13 patients were ophthalmologically evaluated. The age of onset varied after five years of age to early twenties, and the BCVA ranged from 20/60 to no light perception. All the studied patients were noticed to have nystagmus of either a pendular or jerk waveform, photophobia, The clinical characteristics of the 13 index patients are summarized in Table 1.

FA#	PT#	Gen- der	Age	Pt status	Pheno.	Age of onset (yrs.)	Nystagmus	Anterior segment	Fundus	ERG
F1	P1	М	23	Index	STGD	10	Yes	NL	Pale optic disc, retinal vessels attenuation with well demarcated macular coloboma.	Extinguished
F2	P2	F	34	Index	STGD	20	Yes	Cataract	Pale optic disc, retinal vessels attenuation with well demarcated macular coloboma.	Extinguished
F3	Р3	F	26	Index	CRD	5	Yes	NL	Pale optic disc, retinal vessels attenuation and intraretinal bone spicules	(Photobic > Scoto- pic ) dysfunction
	P4	F	15	Sister	CRD	7	Yes	NL	Pale optic disc, retinal vessels attenuation with no sign of RPE pigmentation	(Photobic > Scoto- pic ) dysfunction
	P5	F	9	Sister	CRD	5	Yes	NL	Pale optic disc, retinal vessels at- tenuation with RPE well-defined, oval-shaped retinal pigment epi- thelial defects at the foveal are	(Photobic > Scoto- pic ) dysfunction
	P6	F	8	Sister	CRD	6	Yes	NL	Pale optic disc, retinal vessels attenuation with general RPE mottling	(Photobic > Scoto- pic ) dysfunction

31

**Citation:** Basamat AlMoallem. "Distinct Phenotypic Patterns of *ABCA4* Associated Retinal Disease in a Unique Saudi Consanguineous Cohort". *Acta Scientific Clinical Case Reports* 4.11 (2023): 30-35.

F4	P7	М	15	Index	CRD	7	Yes	NL	Pale optic disc, retinal vessels attenuation with no sign of RPE pigmentation	(Photobic > Scoto- pic ) dysfunction
	P8	F	10	Sister	CRD	5	Yes	NL	Pale optic disc, retinal vessels attenuation with no sign of RPE pigmentation	(Photobic > Scoto- pic ) dysfunction
F5	Р9	F	13	Index	RP	8	Yes	NL	Pale optic disc, retinal vessels attenuation and intraretinal bone spicules	Non-recordable responses
	P10	М	8	Brother	RP	5	Yes	NL	Pale optic disc, retinal vessels attenuation and intraretinal bone spicules	Non-recordable responses
F6	P11	М	40	Index	RP	18	Yes	Cataract	Pale optic disc, retinal vessels attenuation and intraretinal bone spicules	Non-recordable responses
F7	P12	М	22	Index	RP	20	Yes	NL	Pale optic disc, retinal vessels attenuation and intraretinal bone spiculesNon-recordal responses	
F8	P13	М	16	Index	RP	5	Yes	NL	Pale optic disc, retinal vessels attenuation and intraretinal bone spicules	Non-recordable responses

Table 1: Clinical features of thirteen patients from eight unrelated consanguineous families with ABCA4-associated diseases.Abbreviations used: FA#: family number; PT#: patient number; M: male; F: female; BCVA: best-corrected visual acuity;ERG: electroretinogram; OU: both eyes; M: male;; NA: not available; NL: normal limit; NR: non-recordable; SN: subnormal;STGD: Stargardt disease; RP: retinitis pigmentosa; CRD: cone-rod dystrophy.

Overall, the phenotypes on retinal imaging represent the typical known phenotypic heterogeneity of AAD, where the retina may appear normal initially, while later, a variety of abnormalities may develop either isolated or combined with vascular attenuation, intraretinal pigment migration, macular coloboma and optic disc abnormalities. Several phenotype such as in 2/13 (15%), Retinitis Pigmentosa in 5/13 (39%) and Cone- Rod dystrophy in 6/13 (46%) and) were observed in the studied patients. Noteworthy, there were a clear phenotypic variability in the fundus photos within affected patients in the same family (Figure 1).

Concerning the ERG data both rod (scotopic) and cone (photopic) responses were abnormal in all patients, ranging between extinguished to non-recordable responses.

#### Novel and known variants in ABCA4

Overall, eight distinct biallelic *ABCA4* mutations were found in all the studied 13 patients (100%). A summary of all variants identified Table 2. All of them the ACMG criteria for (likely) pathogenicity.

#### Discussion

This genetic and phenotypic study characterizes thirteen patients from eight unrelated consanguineous Saudi families; simplex (n = 5) and complex (n = 8).



32

**Figure 1:** Retinal imaging of four affected siblings with ABCA4associated diseases in Family 3.

Color Fundus of right eye (OD) and Left eye(OS) displaying; (patient 3): pale optic disc, retinal vessels attenuation and intraretinal bone spicules, (patient 4) pale optic disc, retinal vessels attenuation with no sign of RPE pigmentation; (Patient 5) pale optic disc, retinal vessels attenuation with RPE well-defined, oval-shaped retinal pigment epithelial defects at the foveal are; (Patient 6) pale optic disc, retinal vessels attenuation with general RPE mottling.

**Citation:** Basamat AlMoallem. "Distinct Phenotypic Patterns of *ABCA4* Associated Retinal Disease in a Unique Saudi Consanguineous Cohort". *Acta Scientific Clinical Case Reports* 4.11 (2023): 30-35.

				1					
FA#	PT#	Gender	Pt status	Pheno.	Mutation		Туре	ACMG	Reference
F1	P1	М	Index	STGD	<i>ABCA4:</i> NM_000350: c.4739T>C:p.Leu1580Ser	НОМ	Missense	4	Known [10]
F2	P2	F	Index	STGD	ABCA4:NM_000350:c.913C>T:p.Q305*	HOM	Nonsense	5	This study
F3	P3	F	Index	CRD	ABCA4:NM_000350:c.G4867C:p.G1623R	HOM	Missense	4	This study
	P4	F	Sister	CRD		HOM	Missense		
	P5	F	Sister	CRD		HOM	Missense		
	P6	F	Sister	CRD		НОМ	Missense		
F4	P7	М	Index	CRD	ABCA4:NM_000350:c.1633_1634dupGAAA:p.		Frame-	5	This study
	P8	F	Sister	CRD	Asn545Lysfs*24	НОМ	shift		
F5	P9	F	Index	RP	ABCA4:NM_000350: c.4316G>A; P.Gly1439Asp	HOM	Missense	4	Known [10]
	P10	М	Brother	RP		НОМ			
F6	P11	М	Index	RP	ABCA4:NM_000350:c.4852T>A:p.Trp1618Arg	HOM	Missense	5	This study
F7	P12	М	Index	RP	<i>ABCA4:</i> NM_000350: c.5391_5392del:p.Ala1798*	НОМ	Nonsense	5	This study
F8	P13	М	Index	RP	ABCA4:NM_000350: c.5460+1G>A	НОМ	Splicing	4	Known [11]

**Table 2:** Variant assessment of the identified ABCA4 mutations in patients with ABCA4-associated diseases.Abbreviations used: FA#: family number; PT#: patient number; M: male; F: female; Geno: genotype; HOM.: STGD: Stargardt disease; RP:retinitis pigmentosa; CRD: cone-rod dystrophy; Geno.: genotype; HOM: homozygous;; Class 4: likely pathogenic and Class 5: pathogenic.

In our study, biallelic pathogenic variants in eight distinct ABCA4 biallelic mutations were identified in all the studied patients (100%), five of which are novel supporting autosomal recessive inheritance.

Other genomic studies of Saudi cohorts have reported a clinical detection rate of 63% but when solely considering autosomal recessive cases this percentage steeply increases to 93%, with almost exclusively homozygous pathogenic variants being identified which emphasizes the advantage of studying consanguineous families [8].

Herein, a clinical diagnosis of AAD-STGD was made mainly in 2/13 (15%). On the other hand, the majority of the studied families exhibited the diagnosis of either AAD-RP in 5/13 (39%) and AAD-CRD in 6/13 (46%) respectively.

Noteworthy, in family 3, with four affected siblings, despite the confirmed genotype with a novel missense *ABCA4*:NM\_000350:c. G4867C:p.G1623R, distinct clinical phenotypes where observed in their color fundus photos ranging between no sign of RPE pigmentation to intraretinal bone spicules with variable range of macular defects. Thus, the dysfunction of *ABCA4* could give rise to a variety of retinal phenotypes classified mostly according to prognostic

and genotypic presdpostion [9]. Thus, understanding the genotype-phenotype correlation will improves the clinical management and pave the gate to understand the natural history of the disease pathogenesis.

33

In conclusion, we took advantage of studying a highly consanguineous families and be able to solve them al. Our study represents a unique cohort of patients originating from consanguineous families with variable retinal phenotypes caused by known and novel recessive *ABCA4* mutations. This study expands the phenotypic – genotypic spectrum of AAD and provides an accurate genetic diagnosis that paves the gates for the opportunity of gene-based therapies in AAD within the scope of precision medicine.

#### Funding

None.

#### **Financial Disclosures**

No financial disclosures.

#### **Other Acknowledgments**

Patients and families for their valuable continuation in this study.

## **Bibliography**

- 1. Katis NI., *et al.* "Viruses of the genus Allium in the Mediterranean region". *Advances in Virus Research* 84 (2012): 163-208.
- Wylie SJ., *et al.* "Phylogenetic analysis of allexiviruses identified on garlic from Australia". *Australasian Plant Disease Notes* 7 (2012): 23-27.
- 3. Torino IC. "Filamentous viruses of garlic in Argentina". *Plant Disease* 76 (1992): 594-596.
- Melo Filho PdeA., *et al.* "Detection of three Allexivirus species infecting garlic in Brazil". *Pesquisa Agropecuária Brasileira* 39 (2004): 735-740.
- Kang SG., *et al.* "Allexivirus transmitted by eriophyid mites in garlic plants". *Journal of Microbiology and Biotechnology* 17.11 (2007): 1833-1840.
- Song SI., et al. "Molecular characterization of the garlic virus X genome". Journal of General Virology 79.Pt 1 (1998): 155-159.
- 7. Sumi S., *et al.* "Complete nucleotide sequences of garlic viruses A and C, members of the newly ratified genus Allexivirus". *Archives of Virology* 144 (1999): 1819-1826.
- 8. Kritzman A., *et al.* "Distribution and Transmission of Iris yellow spot virus". *Plant Disease* 85 (2001): 838-842.
- 9. Bag S., *et al.* "Iris yellow spot virus (Tospovirus: Bunyaviridae): From obscurity to research priority". *Molecular Plant Pathology* 16 (2015): 224-237.
- Abraham AD., *et al.* "Molecular identification, incidence and phylogenetic analysis of seven viruses infecting garlic in Ethiopia". *European Journal of Plant Pathology* 155 (2019): 181-191.
- 11. Ayed C., *et al.* "First Report of Garlic virus A, Garlic virus B, and Garlic virus C on Garlic (Allium sativum) in Tunisia". *Plant Disease* 106 (2022): 1312.
- 12. Hamed K., *et al.* "First Report of Garlic common latent virus Infecting Garlic in Sudan". *Plant Disease* 97 (2013): 562.
- El-Wahab AEA. "Aphid-Transmission Efficiency of Two Main Viruses on Garlic in Egypt, Onion Yellow Dwarf Virus (OYDV-G) and Leek Yellow Stripe Virus (LYSV-G), in: *Academic Journal* of Entomology 2.1 (2009): 40-42.

- Hafez EE., *et al.* "First Report of Iris yellow spot virus Infection of Garlic and Egyptian Leek in Egypt". *Plant Disease* 96 (2012): 594.
- Mahmoud SYM., *et al.* "Identification of Onion yellow dwarf potyvirus as one of the major viruses infecting garlic in Egypt". *American-Eurasian Journal of Agricultural and Environmental Sciences* 2 (2007): 746-755.
- Hegazy HH., et al. "Evaluation the Growth Performance of Egyptian Garlic Landraces and In vitro Synseeds of Bulblets Formation". Alexandria Journal of Agricultural Sciences 63 (2018): 339-352.
- 17. Cremer J., *et al.* "Detection and Distribution of Viruses Infecting Garlic Crops in Australia". *Plants (Basel)* 10 (2021): 1013.
- FajardoTV., *et al.* "Garlic viral complex: identification of potyviruses and carlavirus in central Brazil". *Fitopatologia Brasileira* 26 (2001): 619-626.
- 19. Wylie SJ., *et al.* "The global trade in fresh produce and the vagility of plant viruses: a case study in garlic". *PLoS One* 9 (204): e105044.
- 20. Cafrune EE., *et al.* "Effect of Two Allexivirus Isolates on Garlic Yield". *Plant Disease* 90 (2006): 898-904.
- 21. Conci VC., *et al.* "Yield losses associated with virus-infected garlic plants during five successive years". *Plant Disease* 87 (2003): 1411-1415.
- 22. Lunello P., *et al.* "Yield Loss in Garlic Caused by Leek yellow stripe virus Argentinean Isolate". *Plant Disease* 91 (2007): 153-158.
- 23. Chen J., *et al.* "A universal PCR primer to detect members of the Potyviridae and its use to examine the taxonomic status of several members of the family". *Archives of Virology* 146 (2001): 757-766.
- Chu FH., *et al.* "Completion of the Genome Sequence of Watermelon silver mottle virus and Utilization of Degenerate Primers for Detecting Tospoviruses in Five Serogroups". *Phytopathology* 91 (2001): 361-368.
- 25. Hassani-Mehraban A., *et al.* "Generic RT-PCR tests for detection and identification of tospoviruses". *Journal of Virological Methods* 233 (2016): 89-96.

- 26. Chen J., *et al.* "Detection and classification of allexiviruses from garlic in China". *Archives of Virology* 149 (2004): 435-445.
- Gibbs A and Mackenzie A. "A primer pair for amplifying part of the genome of all potyvirids by RT-PCR". *Journal of Virological Methods* 63 (1997): 9-16.
- 28. Zheng L., *et al.* "A novel pair of universal primers for the detection of potyviruses". *Plant Pathology* 59 (2010): 211-220.
- Chodorska M., *et al.* "Occurrence of the viruses belonging to the Allexivirus genus on garlic plants in Poland". *Progress in Plant Protection* 53 (2013): 2015-2041.
- 30. Gieck SL., *et al.* "First Report of Garlic virus B and Garlic virus D in Garlic in the Pacific Northwest". *Plant Disease* 93 (2009): 431.
- Hall T. "BioEdit. Biological sequence alignment editor for Win (2005): 95.
- Tamura K., *et al.* "MEGA11: Molecular Evolutionary Genetics Analysis Version 11". *Molecular Biology and Evolution* 38 (2021): 3022-3027.
- Lot H., et al. "Effects of Onion Yellow Dwarf and Leek Yellow Stripe Viruses on Symptomatology and Yield Loss of Three French Garlic Cultivars". *Plant Disease* 82 (1998): 1381-1385.
- Perotto MC., *et al.* "The effect of additional viral infections on garlic plants initially infected with Allexiviruses". *European Journal of Plant Pathology* 126 (2010): 489-495.
- 35. Conci VC., *et al.* "The distribution of garlic viruses in leaves and bulbs during the first year of infection". *Journal of Phytopathology* 158 (2010): 186-193.
- 36. Verbeek M., *et al.* "Efficiency of eradication of four viruses from garlic (Allium sativum) by meristem-tip culture". *European Journal of Plant Pathology* 101 (1995): 231-239.
- Bereda M., *et al.* "Occurrence and phylogenetic analysis of allexiviruses identified on garlic from China, Spain and Poland commercially available on the Polish retail market". *European Journal of Plant Pathology* 149 (2017): 227-237.
- Dabrowska E., *et al.* "Transmission of Garlic virus B, Garlic virus C, Garlic virus D and Garlic virus X by Aceria tulipae (Keifer) in leek". *European Journal of Plant Pathology* 157 (2020): 215-222.

- Taglienti A., *et al.* "Molecular Identification of Allexiviruses in a complex mixture of garlic viruses in Latium". *(Central Italy. European Journal of Plant Pathology* 150 (2018): 797-801.
- 40. Mohammed HS., *et al.* "Occurrence and phylogenetic analysis of potyviruses, carlaviruses and allexiviruses in garlic in Sudan". *Journal of Phytopathology* 161 (2013): 642-650.
- 41. Dovas CI., *et al.* "Comparisons of methods for virus detection in Allium spp". *Phytopathology* 149 (2001): 731-737.
- Dovas I., *et al.* "Incidence of viruses infecting Allium spp. in Greece". *European Journal of Plant Pathology* 107 (2001): 677-684.
- Dovas I and Vovlas C. "Viruses infecting alliums spp. in southern Italy". *Journal of Plant Pathology* 85, 135.
- 44. Fayad-André M de S., *et al.* "Spread of viruses in garlic fields cultivated under different agricultural production systems in Brazil". *Tropical Plant Pathology* 36 (2011): 341-349.
- 45. Parrano L., *et al.* "Characterization of viruses associated with garlic plants propagated from different reproductive tissues from Italy and other geographic regions". *Phytopathologia Mediterranea* (2012): 549-565.
- 46. Mansouri F and Ryšánek P. "Allexivirus: review and perspectives". *Phytopathologia Mediterranea* 60.3 (2021): 389-402.
- Koo BJ., *et al.* "Survey of garlic virus disease and phylogenetic characterization of garlic viruses of the genus Allexivirus isolated in Korea". *The Plant Pathology Journal* 18 (2002): 237-243.

35