- Group 3: Glycosyltransferases (GTFs), which are found in the
 cell periplasmic membrane, are included in the third group
 of enzymes. GTFs transport the sugar nucleotide UDP-Glu or
 UDP-Gal to the repeating unit linked to the glycosyl carrier
 lipid, isoprenoid alcohol [24].
- Group 4: Polysaccharides modified by enzymatic activities such as acetylation, acylation, sulphation, and methylation are exported to the extracellular surface with the help of hydrophobic enzymes such as flippase, permease, or ABC transporters at the fourth stage of synthesis [24].

Bacterial cells primarily use four general strategies for EPS biosynthesis, which are:

Pathway dependent on Wzx/Wzy

First synthesis of nucleotide sugars, assembly of repeating units, polymerization, and export are all part of this route [36]. Activated sugar molecules are transported to and connected with the carrier lipid molecule isoprenoid alcohol in the first stage. GTFs then join other sugar molecules together to create repeating units. These repeating units are now transported across the cytoplasmic membrane by the Wzx flippase [9,15,33,36]. Translocated oligosaccharide undergoes several enzymatic changes, such as methylation, acylation, and others, before being polymerized into polysaccharide by Wzy protein [15]. ABC transporters deliver the formed polysaccharide to the cell surface [9].

Pathway dependent on ABC transporters

The process involved all three members of the ABC Transporter-Dependent Periplasmic protein, Polysaccharide co-polymerase (PCP), and Outer Membrane Polysaccharide Export (OPX) families [8]. Glycosyltransferase is employed in this process to produce polysaccharides before they are exported from the cytoplasm via the tripartite efflux pump complex. When ATP binding and depolymerization (hydrolysis) action at the nucleotide binding domain (NBD) occurs, conformation changes in the membrane heterooligomeric complex of the OPX protein and PCP occur. This pathway is mainly involved in capsular polysaccharide [8].

Pathway dependent on synthase

The polymer products of the Synthase-Dependent Pathway are homopolysaccharides such as cellulose [47,49]. This pathway can be governed with or without the presence of a lipid acceptor molecule. A single synthase protein and membrane-localized GTFs

also manage the polymerization of the EPS precursor and eventual transport of the molecule in this pathway [50]. A complete polymer strand of repeating units is transported across the cell membrane by the flippase enzyme. In comparison to other pathways, this one is largely independent of central carbon metabolism.

Extracellular biosynthesis by sucrase protein

Outside the cellular outer membrane, the extracellular sucrase enzyme transforms sucrose into monomeric units [47]. Dextran, alternant, and Levan are examples of extracellularly generated EPSs. *Leuconostoc mesenteroides* produced dextran with a molecular weight ranging from 15 to 20,000 kDa. According to the reaction below, glycosyltransferase dextransucrase transfers glucose from sucrose to the reducing end of a developing dextran chain.

Sucrose — Dextran + D-fructose

Application of exopolysaccharides

Food industry

Dextran is the first industrial polysaccharide delivered by LAB like Leu. mesenteroides. Dextran was first identified in sugar cane and beet syrups, where it was discovered to be a good thickening and gelating agent [29]. It acts as a gelling agent in gum and jelly candies. It acts as a crystallization inhibitor in ice cream and improves the body texture and mouthfeel of pudding combinations. Dextran is also used in several chromatography stationary phases and as a blood plasma extender [29]. Xanthan gum, which is produced by the plant pathogen Xanthomonas campestris, has been termed a "benchmark" product because of its widespread use in both food and non-food applications [44], including dairy by-products, drinks, confectionery, dressing, bakery products, syrups, and pet foods. Because of the high conversion of a substrate (glucose) to polymer (60-70%), xanthan synthesis is reasonably affordable [44]. This polymer demonstrates a high viscosity at low concentrations in solution and strong pseudo-plasticity and is steady over a wide range of pH, temperature, and ionic strength. Fructose oligosaccharides (FOS) are frequently utilized in the food industry because they have a low sweetness relative to sucrose, are caloriefree, and are noncariogenic [29]. Inulin and FOS are commonly employed in food applications due to their prebiotic characteristics. Fructose-based polymers can be digested by gut microflora, resulting in improved intestinal flora and increased mineral absorption [29]. Levan derived from L. sanfranciscensis LTH 2590 also has

prebiotic properties, as demonstrated by numerous *in vitro* experiments [19]. Fructans play an important role in tolerance to cellular stress in plants by stabilizing membranes [28].

The fitness awareness of consumers generated additional demand, especially for low-fat or fat-free dairy products. However, since milk fat contributes to the development of the taste and body texture of dairy products, the removal of this milk fat leads to structural and functional defects in fermented dairy products. The main problems with low-fat yogurt and dahi are a lack of taste and texture [14]. EPS produced by LAB acts as a thickener is used as a natural, appropriate, and superior substitute for a variety of additives. Instead of chemical additives, these cultures meet consumer requirements for products [18], reduce the total solids required without affecting textural characteristics [48]. Low-fat dahi made using various EPS producing cultures of L. Lactis subsp. Lactis PM23, S. thermophilus ST and L. Lactis NCDC 191 are more acceptable [4]. Dextran is obtained from Leuconostoc mesenteroides with application in baking improvers. A study provides evidence that EPS effectively enhances the rheological characteristics of the dough and the quality of the bread [6]. The in-situ production of EPS from sucrose resulted in the formation of other metabolites like mannitol, glucose, and acetate, all of which can help improve bread quality [20].

Pharmaceutical industry and health aspects

Lactic acid bacteria have become increasingly popular as probiotics in recent years. The ability of the LAB probiotic strain to tolerate acid and bile, produce antimicrobial chemicals against pathogenic and cariogenic bacteria and attach and colonize human intestinal mucosa. The capsular polysaccharides might facilitate the adherence of bacteria to biological surfaces, so they stimulate the colonization of different microhabitats. Similarly, Leuconostoc mesenteroides-produced dextran has been employed to make one of the most effective plasma substitutes for usage in shock and blood loss [41]. Glycosaminoglycan heparin, the medicine of choice for treating thromboembolic diseases, has been linked to a lack of effectiveness in antithrombin deficient patients, with side effects as bleeding and thrombocytopenia. As a result, sulphated forms of alginate have been proposed as a viable alternative with increased activity. Anticoagulant, antithrombotic, anti-atherosclerotic, antiangiogenesis, anti-metastatic, and anti-inflammatory are some of the other therapeutic properties related to sulphated forms of alginate [10]. Xanthan and sulphated dextran are applied as antiviral and anticancer tools. Fucopol is recognized as a substance having the potential to be used in anticancer, anti-inflammatory, and immune-enhancer medications due to the high fucose concentration in some EPS [12]. Fungal polysaccharides have traditionally been used to scavenge and treat a wide variety of diseases such as infectious diseases, cancer, and other autoimmune diseases. A water-soluble Morchellaconica polysaccharide (MCP) controls nitric oxide formation in macrophages and increases splenocyte proliferation and acts as a powerful immunomodulatory agent [43]. The unreasonable use of antibiotics leads to more and more drug-resistant microorganisms, which ultimately lead to incurable diseases. It was recently discovered that immunomodulators are a potential substitute for antibiotics. Polysaccharides derived from microorganisms are the main factor in macrophage stimulation to induce the immune system's toll-like receptors [23].

Bioremediation

Environmental contamination has created various social difficulties throughout this period of urbanization. The widespread use of chemicals such as solvents, herbicides, insecticides, and other industrial compounds has contaminated soil, air, rivers, oceans, waste streams, and numerous locations. These environmental pollutants harm a huge number of creatures, causing them to become trapped in food chains. Bioremediation, a biological method of utilizing microbes and plants, can help to the removal of hazardous substances from contaminated sites [37-39].

The use of EPS-producing microbes in the treatment of mining-related environmental effluents is a growing field of biotechnology [16]. The probable role of EPS in the removal of heavy metals from the environment is due to their ability to bind metal ions from solutions. A prominent group of sulfate-reducing bacteria (SRB) is generally located in metal-contaminated wastewaters [42]. In the anaerobic condition, these groups of bacteria are degraded of numerous organic contaminants and the precipitation of heavy metals from wastewater. Other bacteria demonstrating biosorption of toxic heavy metals in bioremediation procedures contain *Enterobacter* and *Pseudomonas* species [40,42]. Fungal-bacterial biofilms (FBBs) were employed for the first time to extract nickel from wastewater. Hexavalent chromium bioremediation in wastewater using FBB is also a unique approach. A study found that established FBBs, glass-wool-attached bacterial biofilms, and their monocul-

tures were effective in removing hexavalent chromium. EPS separated from *Zoogloea* spp. and *Aspergillus niger* support in the degradation of pyrene in contaminated soils [17]. Some EPS-producing bacteria, such as Bacillus cereus possess the capability of biocorrosion of stainless steel and are thus utilized in bioremediation to remove unwanted steel compounds in stainless steel companies [5]. *Sphingomonads* are unique in that they have multiple large pleat-like structures on their cell surface, as well as an extraordinary metabolic capacity to degrade a variety of rigid environmental pollutants, particularly xenobiotics like dioxin, biphenyl, and bisphenol, and the ability to produce valuable biopolymers [1].

The petroleum industry

The petroleum industry uses xanthan gum as a bacterial EPS in oil drilling, fracturing, and pipeline cleaning [1], and it's also useful as a drilling fluid additive because of its salt tolerance and resistance to temperature degradation [46]. Microbial Enhanced Oil Recovery (MEOR) is the process of using microorganisms to extract additional oil from existing reservoirs, hence increasing the petroleum production of an oil reservoir. Select natural microorganisms are introduced into oil reservoirs in this manner, resulting in harmless by-products such as slippery natural compounds or gases, all of which help in the extraction of oil from the well. These techniques help to assemble the oil and facilitate oil movement, allowing for a greater amount of oil to be recovered from the well. Genetically engineered *Enterobacter cloacae* are used in MEOR [1].

Agriculture

The growth of EPS-producing bacteria in the rhizosphere of cultivated plants can improve soil fertility and productivity. Meanwhile, EPS-producing bacteria are present within the roots and surrounding soil, increasing the movement of water and nutrients via the plant roots. The PGPR effect could be linked to very high yields in shoot and root growth of cultivated plants that were produced in a salty environment due to an EPS-forming bacterial inoculation because nutrient uptake and utilization is the limiting method for crop growth and yield in later growth stages [2]. In the terrestrial environment, *Azotobacter* EPS is important for ecosystem function because it regulates nutrient cycle processes, which are necessary for soil productivity, as well as biotic and abiotic pressures in the soil ecosystem [1,36].

Exopolysaccharides are employed as biosurfactants in some cases. Many rhizosphere and plant-associated bacteria generate biosurfactants which are low molecular weight surface-active chemicals, and are important for motility, signal transmission, and biofilm formation, indicating that EPS regulates plant-microbe communication. EPS can be utilized to improve agricultural soil qualities on a wide scale through soil remediation. These biomolecules have the potential to someday replace the hazardous surfactants currently utilized in the multibillion-dollar pesticide industries [34]. The outer surface of rhizobia is made up of complex polysaccharides including lipopolysaccharides (LPS), capsular polysaccharides (CPS), and extracellular polysaccharides (EPS). They can create associations with legumes such as Trifolium, Pisum, Vicia, and Medicago spp and induce the formation of specific organs on roots and stems called nodules, in which atmospheric nitrogen is converted to ammonia by the nitrogenase enzyme complex [1]. The exopolysaccharides gellan and curdlan have agricultural potential because they serve as a soil improver, which improves the cultivated soil's water retention capacity [26].

Textile industry

Because of its viscosity, stabilizer, and cross-linking capabilities with fabrics, EPS is utilized as a binding agent with color dyes or hydrogel in the textile industry. One of the unique applications of EPS is smart fabrics. Hydrogels based on biopolymers exhibit specific physical properties of swelling and shrinkage that are regulated by external factors such as pH, temperature, solvent, electric field, light, stress, ionic strength, and other external chemical stimuli, among others [21]. Biopolymers like chitosan are combined with synthetic polymers to make a hydrogel that can be successfully implanted on fabric surfaces for smart textile applications. At appropriate temperatures, these fabrics are used as deodorant release agents. As a result, they're used in fabric aroma finishing. With a change in external temperature or pH, the polymer beta-Cyclodextrin can release it. These fabrics can also change color in response to changes in external temperature by including thermochromic elements such as cholesterins [1].

Conclusion

Bacterial-produced exopolysaccharides have a wide range of activities and are not restricted by taxa. The monomeric compositions, linking bonds, and associated conjugates demonstrate some of this complexity, while the functions can be categorized into intrinsic and applied. The essential functions in human use, including morphological, structural, and defensive roles, are evident; Medical, cosmetic, pharmaceutical, dairy and other industrial and environmental items are just a few examples. For microbial EPS production, understanding the biosynthetic mechanism is an important topic for optimizing EPS production yields, improving product quality and properties, and also for designing novel strains. Since most bacterial EPS with unique properties have expensive production costs and economic hurdles to overcome, this valuable biosynthetic information is also important to reduce these costs. Since microbial biopolymer biosynthesis is the result of a complex system of many metabolic processes, system-based approaches to control and optimize production are needed to improve the previously reported yields.

Acknowledgments

We are grateful to the Department of Microbiology and Biotechnology, University School of Sciences, Gujarat University, Gujarat, India, for providing us with the necessary resources to finish this task.

Conflicts of Interest

We declare that here are no conflicts of interest.

Bibliography

- Andhare P., et al. "Microbial exopolysaccharides: advances in applications and future prospects". Biotechnology 3 (2014): 25.
- 2. Ashraf M., *et al.* "Inoculating wheat seedlings with exopolysaccharide–producing bacteria restricts sodium uptake and stimulates plant growth under salt stress". *Biology and Fertility of Soils* 40.3 (2004): 157-162.
- 3. Badel S., *et al.* "New perspectives for Lactobacilli exopolysac-charides". *Biotechnology Advances* 29.1 (2011): 54-66.
- Behare P., et al. "Exopolysaccharide-producing mesophilic lactic cultures for preparation of fat-free Dahi - an Indian fermented milk". The Journal of Dairy Research 76 (2009): 90-97.
- Bragadeeswaran S., et al. "Exopolysaccharide production by Bacillus cereus GU812900, a fouling marine bacterium". African Journal of Microbiology Research 5.24 (2011): 4124-4132.

- Brandt MJ., et al. "Effect of an exopolysaccharide produced by Lactobacillus sanfranciscensis LTH1729 on dough and bread quality". Sourdough from Fundamentals to Applications (2003): 80.
- 7. Casillo A., *et al.* "Exopolysaccharides from marine and marine extremophilic bacteria: structures, properties, ecological roles and applications". *Marine Drugs* 16.2 (2018): 69.
- Cuthbertson L., et al. "Pivotal roles of the outer membrane polysaccharide export and polysaccharide copolymerase protein families in export of extracellular polysaccharides in gram-negative bacteria". Microbiology and Molecular Biology Reviews 73.1 (2009): 155-177.
- 9. De Vuyst L., *et al.* "Recent developments in the biosynthesis and applications of heteropolysaccharides from lactic acid bacteria". *The International Dairy Journal* 11 (2001): 687-707.
- DeAngelis PL and White CL. "Identification and Molecular Cloning of a Heparosan Synthase from Pasteurellamultocida Type D". *Journal of Biological Chemistry* 277.9 (2002): 7209-7213.
- 11. Dwivedi M. "Exopolysaccharide (EPS) producing isolates from sugarcane field soil and antibacterial activity of extracted EPSs". *Acta Scientific Microbiology* 1 (2018): 06-13.
- 12. Freitas F., *et al.* "Advances in bacterial exopolysaccharides: From production to biotechnological applications". *Trend in Biotechnology* 29.8 (2011): 388-398.
- 13. Gupta P and Diwan B. "Bacterial exopolysaccharide mediated heavy metal removal: a review on biosynthesis, mechanism and remediation strategies". *Biotechnology Reports* 13 (2017): 58-71.
- 14. Guven M., *et al.* "The effect of inulin as a fat replacer on the quality of set-type low-fat yogurt manufacture". *International Journal of Dairy Technology* 58 (2005): 180-184.
- 15. Islam ST and Lam JS. "Synthesis of bacterial polysaccharides via the Wzx/Wzy-dependent pathway". *Canadian Journal of Microbiology* 60.11 (2014): 697-716.
- 16. Iyer A., et al. "Emulsifying properties of a marine bacterial exopolysaccharide". Enzyme and Microbial Technology 38.1-2 (2006): 220-222.
- 17. Jia C., *et al.* "Degradation of pyrene in soils by extracellular polymeric substances (EPS) extracted from liquid cultures". *Process Biochemistry* 46.8 (2011): 1627-1631.

- 18. Jolly L., et al. "Exploiting exopolysaccharides from lactic acid bacteria". Lactic Acid Bacteria: Genetics, Metabolism and Applications (2002): 367-374.
- Korakli M., et al. "Metabolism by bifidobacteria and lactic acid bacteria of polysaccharides from wheat and rye, and exopolysaccharides produced by Lactobacillus sanfranciscensis". Journal of Applied Microbiology 2 (2002): 958-965.
- Korakli M., et al. "Sucrose metabolism and exopolysaccharide production in wheat and rye sourdoughs by Lactobacillus sanfranciscensis". Journal of Agricultural and Food Chemistry 49.11 (2001): 5194-5200.
- 21. Kulkarni A., *et al.* "Microgel-based surface modifying system for stimuli-responsive functional finishing of cotton". *Carbohydrate Polymers* 82.4 (2010): 1306-1314.
- 22. Li J and Wang N. "The gpsX gene encoding a glycosyltransferase is important for polysaccharide production and required for full virulence in Xanthomonas citri subsp". citri. *Bmc Microbiology* 12.1 (2012): 1-16.
- 23. Lin MH., *et al.* "A novel exopolysaccharide from the biofilm of Thermus aquaticus YT–1 induces the immune response through toll– like receptor 2". *Journal of Biological Chemistry* 286.20 (2011): 17736-17745.
- 24. Mishra A and Jha B. "Microbial exopolysaccharides". *The Pro-karyotes* 4 (2013): 179-192.
- 25. MohdNadzir M., *et al.* "Biomedical applications of bacterial exopolysaccharides: A review". *Polymers* 13.4 (2021): 530.
- 26. Morris G and Harding S. "Polysaccharides, microbial". *Applied Microbiology: Industrial* (2009): 482-494.
- 27. Nwodo UU., *et al.* "Bacterial exopolysaccharides: functionality and prospects". *International Journal of Molecular Sciences* 13.11 (2012): 14002-14015.
- 28. Oliver AE., *et al.* "Nondisaccharide-based mechanisms of protection during drying". *Cryobiology* 43 (2001): 151-167.
- 29. Patel A and Prajapat JB. "Food and health applications of exopolysaccharides produced by lactic acid bacteria". *Advances in Dairy Research* (2013): 1-8.
- Pawar ST., et al. "Isolation, screening and optimization of exopolysaccharide producing bacterium from saline soil". Journal of Microbiology and Biotechnology Research 3.3 (2013): 24-31.

- 31. Pérez-Burgos M., *et al*. "Characterization of the exopolysaccharide biosynthesis pathway in Myxococcus xanthus". *Journal of Bacteriology* 202.19 (2020): e00335-20.
- 32. Rana S and Upadhyay LSB. "Microbial exopolysaccharides: Synthesis pathways, types and their commercial applications". *International Journal of Biological Macromolecules* 157 (2020): 577-583.
- 33. Rehm BH. "Bacterial polymers: biosynthesis, modifications and applications". *Nature Reviews Microbiology* 8.8 (2010): 578-592.
- 34. Sachdev DP and Cameotra SS. "Biosurfactants in agriculture". *Applied Microbiology and Biotechnology* 97.3 (2013): 1005-1016.
- 35. Sanalibaba P and Çakmak GA. "Exopolysaccharides production by lactic acid bacteria". *Applied Microbiology: Open Access* 2.1000115 (2016).
- 36. Schmid J., et al. "Bacterial exopolysaccharides: biosynthesis pathways and engineering strategies". Frontiers in Microbiology 6 (2015): 496.
- 37. Sharma S., *et al.* "Biofilm: Used as A Brand-new Technology in Bioremediation". Vidya; A Journal of Gujarat University 16.2 (2021): 9-116.
- 38. Sharma S., et al. "Phytomining of Heavy Metals: A Green Technology to Sustainable Agriculture". International Journal of Innovative Research in Science, Engineering and Technology (IJIR-SET) 10.6 (2021): 7527-7538.
- Sharma S., et al. "Isolation of Heavy Metal Tolerant Rhizobacteria from Zawar Mines Area, Udaipur, Rajasthan, India". Bioscience Biotechnology Research Communication 13.1 (2020): 233-238.
- 40. Sharma Sarita., et al. "Elucidate the Influence of Heavy Metal on Bacterial Growth Isolated from a Mining Location and A Waste Dump: Using their Inducible Mechanism". The Current Trends in Biomedical Engineering and Biosciences 20.2 (2021): 556034.
- 41. Silver RP, *et al.* "The K1 capsular polysaccharide of Escherichia coli". *Reviews of Infectious Diseases* (1988): S282-S286.
- 42. Singha TK. "Microbial extracellular polymeric substances: production, isolation and applications". *IOSR Journal of Pharmacy* 2.2 (2012): 271-281.

- 43. Su CA., et al. "Isolation and characterization of exopolysaccharide with immunomodulatory activity from fermentation broth of Morchellaconica". *DARU Journal of Pharmaceutical Sciences* 21.1 (2013): 1-6.
- 44. Sutherland IW. "Novel and established applications of microbial polysaccharides". *Trends in Biotechnology* 16 (1998): 41-46.
- 45. Sutherland IW. "Biotechnology of microbial exopolysaccharides, No. 9". Cambridge University Press (1990).
- 46. Sutherland IW. "Microbial polysaccharides from Gram-negative bacteria". *International Dairy Journal* 11.9 (2001): 663-674.
- 47. Ua-Arak T., *et al.* "Fermentation pH modulates the size distributions and functional properties of Gluconobacteralbidus TMW 2.1191 levan". *Frontiers in Microbiology* 8 (2017): 807.
- 48. Wacher-Rodarte C., *et al.* "Yogurt production from reconstituted skim milk powders using different polymer and non-polymer forming starter cultures". *The Journal of Dairy Research* 60 (1993): 247-254.
- 49. Wang J., et al. "Characterization and immunomodulatory activity of an exopolysaccharide produced by Lactobacillus plantarum JLK0142 isolated from fermented dairy tofu". International Journal of Biological Macromolecules 115 (2018): 985-993.
- 50. Whitney JC and Howell PL. "Synthase-dependent exopolysaccharide secretion in Gram-negative bacteria". *Trends in Microbiology* 21.2 (2013): 63-72.

Assets from publication with us

- · Prompt Acknowledgement after receiving the article
- Thorough Double blinded peer review
- · Rapid Publication
- Issue of Publication Certificate
- High visibility of your Published work

Website: www.actascientific.com/

Submit Article: www.actascientific.com/submission.php

Email us: editor@actascientific.com Contact us: +91 9182824667